



# Sub-chronic exposure to titanium dioxide and cerium oxide nanoparticles alone or their combination induced hepatotoxicity and neurotoxicity in male rats

Thulfiqar Fawwaz Mutar✉

Department of Environmental Studies, Institute of Graduate Studies and Research, Alexandria University, Alexandria, Egypt

✉Corresponding Author:

Department of Environmental Studies,  
Institute of Graduate Studies and Research,  
Alexandria University, Alexandria,  
Egypt  
E-mail: thulficarfawwaz@gmail.com,  
Tel. +20 – 1100671504

## Article History

Received: 09 September 2017

Accepted: 24 October 2017

Published: July-December 2017

## Citation

Thulfiqar Fawwaz Mutar. Sub-chronic exposure to titanium dioxide and cerium oxide nanoparticles alone or their combination induced hepatotoxicity and neurotoxicity in male rats. *Drug Discovery*, 2017, 11(30), 14-22

## Publication License



© The Author(s) 2017. Open Access. This article is licensed under a Creative Commons Attribution License 4.0 (CC BY 4.0).

## General Note



Article is recommended to print as color version in recycled paper.

## ABSTRACT

Titanium dioxide nanoparticles (TiO<sub>2</sub>NPs) and cerium oxide nanoparticles (CeO<sub>2</sub>NPs) are widely used in many applications fields nowadays and it has concern the attention of scientists in recent years as a result of toxicity. The objective of this study was to detect liver and brain toxicity induced by TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs or their combination in male rats. Animals were administered orally with TiO<sub>2</sub>NPs (100 mg/kg bw) and CeO<sub>2</sub>NPs (50 mg/kg bw) alone or in combination every day for 60 days. TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs or their combination was confirmed by the elevation in tissues inflammatory markers including: (IFN $\gamma$ , TNF- $\alpha$  and IL-6). Moreover, (TAC) was decreased in liver and brain tissues while (NO) was increased. Also, caused decreased (DA) and serotonin in plasma and brain while (ACh) was increased compared to control group. (ALT, AST, AIP and LDH) were increased in plasma and decreased in liver. The data show that administration of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs caused an undesirable effect on most of liver and brain parameters in liver and brain. Also, its combination proved to be even more toxic than each one.

**Key words:** Titanium dioxide nanoparticles; Cerium oxide nanoparticles; Male rats; Hepatotoxicity; Neurotoxicity

**Abbreviations:** IFN $\gamma$ , Interferon gamma; TNF- $\alpha$ , tumor necrosis factor- alpha; IL-6, interleukin-6; TAC, total antioxidant capacity; NO, nitric oxide; DA, dopamine; Ach, Acetylcholine; ALT, Alanine transaminase; AST, Aspartate transaminase; AIP, Alkaline phosphatase; LDH, lactate dehydrogenase; ROS, reactive oxygen species.

## INTRODUCTION

Nanoparticles (NPs) are particle less than 100 nm in at least one critical dimension. Because of their greater surface area display properties not shown by their macroscopic counterparts; this has led to the rapid development of a new field, nanomedicine, which provides potent applications for many purposes such as diagnostic, imaging, therapeutic and drug delivery (De Jong and Borm, 2008). The nanotoxicology based on the acquisition and development of a comprehensive understanding of the relationship between the toxicity of NPs depending on their dose levels and physicochemical properties such as size, shape and configures interactive materials (Paur et al., 2011). NPs are known to induce (ROS) production, leading to an oxidative stress when redox state of the cell is imbalanced (Lin et al., 2008 and Sarkar et al., 2014). Titanium dioxide nanoparticles (TiO<sub>2</sub>NPs) it is used in wide applications due to its chemical and physical characteristics, air purification in biomedical fields, cosmetics industry, sewage purification, food packaging and painting. Because of the wide applications of TiO<sub>2</sub>NPs the potential for human exposure and its release into the environment will be significant, which will allow an increase of the health effects on humans. Previous studies suggest that exposure to TiO<sub>2</sub> NPs leads to many effects on cytotoxicity and genetic toxicity (Gurr et al., 2005 and Sadiq 2012).

Exposure to TiO<sub>2</sub>NPs caused congestion and proliferation of spleen, liver, kidney and brain tissues, with accompanying increases ROS in those tissues. The elevated ROS levels in affected organs lead to lipid peroxidation and proinflammatory cytokines. Because of the accumulation of nanoparticles in organs may exert cytotoxic effects through the induction of oxidative stress (Wang et al., 2011). However, studies have found that TiO<sub>2</sub>NPs were accumulated in the damaged brain and induced apoptosis and oxidative stress in microglia, neurons of mice after sub chronic exposure (Yu et al., 2008 and Wu et al., 2009). Research is still insufficient to assess the impact and toxicity of TiO<sub>2</sub>NPs. Need more evidence, mechanism and detectors to understand the negative effects of TiO<sub>2</sub>NPs and counteract the toxic effects. The possible toxic mechanisms of TiO<sub>2</sub>NPs directly or indirectly were related with generation and accumulation of (ROS) which give rise to an inflammatory response (Jaeger et al., 2012 and Shi et al., 2013). Cerium oxide nanoparticles (CeO<sub>2</sub>NPs) it is widely used applications as a catalyst, ultraviolet absorber and biomedical materials because of the redox activities, can be used as an reducing fuel consumption, greenhouse gases and particle numbers in vehicle exhaust, additive to improve the burning efficiency of fuels (Wang and Lin 2004; Selvan et al., 2009; Lee et al., 2013 and Xu and Qu, 2014). The exposure and distribution of CeO<sub>2</sub>NPs by oral are the most common ways to mice because gastrointestinal region is one of the most important portals of entry for NPs in humans and animals (Hirst et al., 2013). Previous studies which have investigated CeO<sub>2</sub>NPs induced toxicity through different routes of exposure in rats such as inhalation, intra tracheal instillation, and intravenous (iv) administration (Srinivas et al., 2011; Ma et al., 2012 and Tseng et al., 2012). Exposure to CeO<sub>2</sub>NPs after inhalation induced effects in lung and brain could enhance the levels of pro inflammatory cytokine, tumor necrosis factor- alpha (TNF- $\alpha$ ) and interleukin-6 (IL-6) in the cerebellum, and brain stem of atherosclerosis-prone mice (Mohammad et al., 2017; Geraets et al., 2012 and Aalapati et al., 2014). Administration of CeO<sub>2</sub>NPs in monocrotaline induced liver toxicity in rat models results increase hepatic catalase and superoxide dismutase (SOD) activities (Amin et al., 2011). The objective of this study was to investigate the potential toxic effect of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs alone or in combination on liver and brain of male rats through determine the (IFN $\gamma$ ), tumor

necrosis factor-  $\alpha$  (TNF- $\alpha$ ), interleukin-6 (IL-6), acetylcholine (ACh), dopamine (DA), serotonin, total antioxidant capacity (TAC), nitric oxide (NO) and liver enzyme activities.

## MATERIALS AND METHODS

### Chemicals and doses

Titanium dioxide nanoparticles (TiO<sub>2</sub>NPs) <21 nm size and cerium oxide nanoparticles (CeO<sub>2</sub>NPs) < 25 nm size were purchased from Sigma Chemical Company (St. Louis, MO, USA), were dissolved in distilled water. The dose of TiO<sub>2</sub>NPs was (100 mg/kg bw) and was chosen according to Morgan et al. (2017). The dose of CeO<sub>2</sub>NPs was (50 mg/kg bw) and was chosen according to Kumari et al. (2014).

### Animals and experimental groups

Sixty healthy Male Wistar rats weighing  $160 \pm 10$  g, were obtained from the Animal Breeding House of the National Research Centre (NRC), Cairo, Egypt. The housing and management of the animals and the experimental protocols were conducted as stipulated in the Guide for Care and Use of Laboratory Animals (NRC, 1996). The rats were allowed to acclimatize for a week before starting the experiments. Rats were maintained under temperature-controlled. They were fed with standard food and had free access to water. Animals were randomly divided into four groups: a control group and three treatment groups: TiO<sub>2</sub>NPs (100 mg/kg bw; <21 nm), CeO<sub>2</sub>NPs (50 mg/kg bw; <25 nm) and combination of TiO<sub>2</sub>NPs plus CeO<sub>2</sub>NPs, respectively. Rats were orally administered their respective doses every day for 60 consecutive days. With one group assigned to be an untreated control. The experimental work on rats was performed with the approval of the Animal Care & Experimental Committee.

### Blood samples collection

At the end of the experimental period, rats were anesthetized using diethyl ether. Blood samples were taken from the vena cava of rat heart. Tubes were used to compile blood drawn from the heart directly; 1 ml were collected on sodium heparin for hematological studies and placed immediately on ice, the rest of the blood was collected in tubes for coagulation and plasma formation. The collected blood was centrifuged at 860 Xg for 20 min for the separation of plasma and kept at - 80°C until analyses of the tested parameters. The abdominal cavity of each rat was opened where the liver and heart were excised. These tissues were further used for enzymes analyses. The tissues stored in -80°C till assays and homogenized (10%, w/v), separately, in ice-cold sucrose buffer (0.25 M) in a Potter–Elvehjem type homogenizer at 11000 Xg for 20 min at 4 °C for the determination of tested enzyme assays.

### Determination of cytokines, total antioxidant capacity and nitric oxide in liver and brain

IFN $\gamma$ , TNF- $\alpha$  and IL-6 were assayed by using Enzyme-linked Immunosorbent Assay (ELISA) kits in tissue homogenates according to the methods of Shalaby et al. (1985), Hedayati et al. (2001) and Ferguson-Smith et al. (1988), respectively. TAC and the level of NO were assayed in organs homogenates according to the manual instruction of Biodiagnostic Kit, Egypt.

### Determination of liver enzyme activities in plasma and liver

The activities of plasma and liver (AST, ALT, ALP and LDH) were measured with kits from Biosystems S.A (Costa Brava 30, Barcelona, Spain).

### Determination of acetylcholine, Dopamine and serotonin in plasma and brain

The determination of plasma and brain (ACh, DA and serotonin) by Enzyme-linked Immunosorbent Assay (ELISA) using a competitive inhibition enzyme immunoassay technique for the *in vitro* quantitative measurement of kits (Designed by Cloud-Clone Corp. Houston, USA).

### Statistical analysis

Values obtained as means  $\pm$  SE were subjected to for all studied parameters were performed using the general linear model (GLM) produced by Statistical Analysis Systems Institute (SAS, 1998). Duncan's New Multiple Range Test was used to test the significance of the differences between means (Duncan, 1955). Values of  $P < .05$  were considered significant.

## RESULTS

### Effect of titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination on Interferon gamma, tumor necrosis factor- $\alpha$ , interleukin-6, total antioxidant capacity and nitric oxide in liver and brain

As shown in Table 1, the changes in (IFN $\gamma$ , TNF- $\alpha$ , IL-6, TAC and NO levels in liver and brain of male rats. Administration of all the three groups (TiO<sub>2</sub>NPs),(CeO<sub>2</sub>NPs) and their combination led to a significant increase ( $P < 0.05$ ) in IFN $\gamma$ , TNF- $\alpha$ , IL-6 and NO levels while there was significant decrease ( $P < 0.05$ ) in TAC activity compared to the control group. Among the exposed groups, levels were found to be the highest changes in case of combination group, suggesting enhanced increase toxicity potential of combination in comparison to other groups.

### Effect of titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination on Acetylcholine (ACh), Dopamine (DA) and serotonin in plasma and brain

The levels of ACh, DA and serotonin were measured in plasma and brain of male rat's administration daily for 60 days with (TiO<sub>2</sub>NPs),(CeO<sub>2</sub>NPs) and their combination experimental were shown in Table 2. Data showed that treatment with (TiO<sub>2</sub>NPs),(CeO<sub>2</sub>NPs) and their combination caused significant ( $P < 0.05$ ) decrease DA and serotonin in plasma and brain while ACh was significant ( $P < 0.05$ ) increase compared to control group. The effects in the combination were more than the rest of the groups.

### Effect of titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination on the enzymes activities in plasma and liver

The ALT, AST, AIP, and LDH were measured in plasma and liver of male rats with (TiO<sub>2</sub>NPs), (CeO<sub>2</sub>NPs) and their combination. Table 3, Data showed that treatment with TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs and their combination caused significant ( $P < 0.05$ ) increase plasma and decrease liver ALT, AST, AIP and LDH activities compared to control group. While the combination group was significantly ( $P < 0.05$ ) changes compared to either groups.

**Table 1** Mean values  $\pm$  SE of liver and brain levels of interferon gamma, tumor necrosis factor- $\alpha$ , interleukin-6, total antioxidant capacity, and nitric oxide of male rats treated with titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination

Parameter	Experimental groups			
	Control	TiO <sub>2</sub> NPs	CeO <sub>2</sub> NPs	TiO <sub>2</sub> NPs + CeO <sub>2</sub> NPs
<b>Liver</b>				
IFN $\gamma$ (ng /mg tissue)	66.6 $\pm$ 2.38 <sup>c</sup>	129 $\pm$ 3.61 <sup>b</sup>	142 $\pm$ 3.55 <sup>b</sup>	25 $\pm$ 3.58 <sup>a</sup>
TNF- $\alpha$ (ng/gm tissue)	145 $\pm$ 5.66 <sup>d</sup>	326 $\pm$ 5.15 <sup>c</sup>	399 $\pm$ 4.91 <sup>b</sup>	481 $\pm$ 6.83 <sup>a</sup>
IL-6 (ng/gm tissue)	165 $\pm$ 3.2 <sup>d</sup>	215 $\pm$ 3.1 <sup>b</sup>	192 $\pm$ 2.8 <sup>c</sup>	266 $\pm$ 6.1 <sup>a</sup>
TAC ( $\mu$ M/ gm tissue)	2.7 $\pm$ 0.19 <sup>a</sup>	1.87 $\pm$ 0.15 <sup>b</sup>	1.82 $\pm$ 0.13 <sup>b</sup>	1.66 $\pm$ 0.15 <sup>c</sup>
NO ( $\mu$ mole/gm tissue)	0.36 $\pm$ 0.11 <sup>c</sup>	0.58 $\pm$ 0.17 <sup>b</sup>	0.61 $\pm$ 0.14 <sup>b</sup>	0.89 $\pm$ 0.11 <sup>a</sup>
<b>Brain</b>				
IFN $\gamma$ (ng /mg tissue)	87 $\pm$ 2.42 <sup>c</sup>	135 $\pm$ 2.36 <sup>b</sup>	127 $\pm$ 3.12 <sup>b</sup>	177 $\pm$ 4.34 <sup>a</sup>
TNF- $\alpha$ (ng/gm tissue)	121 $\pm$ 5.8 <sup>d</sup>	245 $\pm$ 7.4 <sup>c</sup>	332 $\pm$ 8.3 <sup>b</sup>	418 $\pm$ 6.6 <sup>a</sup>
IL-6 (ng/gm tissue)	144 $\pm$ 4.7 <sup>d</sup>	242 $\pm$ 3.8 <sup>b</sup>	261 $\pm$ 4.1 <sup>c</sup>	398 $\pm$ 3.8 <sup>a</sup>
TAC ( $\mu$ M/ gm tissue)	22.11 $\pm$ 0.33 <sup>a</sup>	15.21 $\pm$ 0.31 <sup>b</sup>	16.10 $\pm$ 0.20	12.44 $\pm$ 0.42 <sup>c</sup>
NO ( $\mu$ mole/gm tissue)	0.47 $\pm$ 0.08 <sup>d</sup>	0.92 $\pm$ 0.18 <sup>b</sup>	0.54 $\pm$ 0.11 <sup>c</sup>	1.32 $\pm$ 0.13 <sup>a</sup>

Mean values within a row not sharing a common superscript letters (a, b, c, d) were significantly different,  $p < 0.05$ .

**Table 2** Mean values  $\pm$  SE of plasma and brain acetylcholine, dopamine and serotonin of male rats treated with titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination

Parameter	Experimental groups			
	Control	TiO <sub>2</sub> NPs	CeO <sub>2</sub> NPs	TiO <sub>2</sub> NPs + CeO <sub>2</sub> NPs
<b>Plasma</b>				
ACh (n mole/ml)	44 $\pm$ 1.6 <sup>d</sup>	69 $\pm$ 2.8 <sup>c</sup>	92 $\pm$ 3.1 <sup>b</sup>	122 $\pm$ 1.9 <sup>a</sup>
DA (ng/ml)	127 $\pm$ 1.8 <sup>a</sup>	87 $\pm$ 2.3 <sup>b</sup>	92 $\pm$ 2.2 <sup>b</sup>	61 $\pm$ 1.7 <sup>c</sup>
Serotonin (ng/ml)	203 $\pm$ 1.7 <sup>a</sup>	155 $\pm$ 2.5 <sup>c</sup>	188 $\pm$ 2.4 <sup>b</sup>	108 $\pm$ 2.6 <sup>d</sup>
<b>Brain</b>				
ACh (n mole/gm tissue)	77 $\pm$ 1.2 <sup>d</sup>	198 $\pm$ 3.1 <sup>c</sup>	221 $\pm$ 8.0 <sup>b</sup>	366 $\pm$ 3.5 <sup>a</sup>
DA (ng /gm tissue)	174 $\pm$ 1.15 <sup>a</sup>	117 $\pm$ 0.88 <sup>b</sup>	112 $\pm$ 2.03 <sup>b</sup>	62 $\pm$ 2.03 <sup>c</sup>
Serotonin (ng /gm tissue)	96 $\pm$ 1.5 <sup>a</sup>	56 $\pm$ 1.5 <sup>c</sup>	67 $\pm$ 1.5 <sup>b</sup>	44 $\pm$ 1.2 <sup>d</sup>

Mean values within a row not sharing a common superscript letters (a, b, c, d) were significantly different,  $p < 0.05$ .

**Table 3** Mean values  $\pm$  SE of plasma and liver alanine transaminase, aspartate transaminase, alkaline phosphatase, and lactate dehydrogenase of male rats treated with titanium dioxide nanoparticles, cerium oxide nanoparticles and their combination

Parameter	Experimental groups			
	Control	TiO <sub>2</sub> NPs	CeO <sub>2</sub> NPs	TiO <sub>2</sub> NPs + CeO <sub>2</sub> NPs
<b>Plasma</b>				
ALT (U/L)	52 $\pm$ 1.4 <sup>c</sup>	95 $\pm$ 1.6 <sup>b</sup>	88 $\pm$ 2.0 <sup>b</sup>	152 $\pm$ 1.9 <sup>a</sup>
AST (U/L)	42 $\pm$ 1.2 <sup>c</sup>	81 $\pm$ 1.7 <sup>b</sup>	73 $\pm$ 1.4 <sup>b</sup>	118 $\pm$ 2.3 <sup>a</sup>
ALP (U/L)	119 $\pm$ 3.51 <sup>c</sup>	159 $\pm$ 2.18 <sup>b</sup>	176 $\pm$ 2.55 <sup>b</sup>	217 $\pm$ 2.19 <sup>a</sup>
LDH (U/L)	1078 $\pm$ 18.6 <sup>c</sup>	1297 $\pm$ 21.5 <sup>b</sup>	1308 $\pm$ 19.9 <sup>b</sup>	1371 $\pm$ 27.7 <sup>a</sup>
<b>Liver</b>				
ALT (U/mg)	366 $\pm$ 4.3 <sup>a</sup>	254 $\pm$ 5.2 <sup>b</sup>	277 $\pm$ 5.1 <sup>b</sup>	200 $\pm$ 3.6 <sup>c</sup>
AST (U/mg)	227 $\pm$ 3.26 <sup>a</sup>	158 $\pm$ 3.24 <sup>b</sup>	169 $\pm$ 3.11 <sup>b</sup>	112 $\pm$ 2.89 <sup>c</sup>
ALP (U/mg)	231 $\pm$ 2.41 <sup>a</sup>	155 $\pm$ 2.36 <sup>b</sup>	148 $\pm$ 3.55 <sup>b</sup>	97 $\pm$ 2.27 <sup>c</sup>
LDH (U/mg)	921 $\pm$ 8.13 <sup>a</sup>	723 $\pm$ 7.38 <sup>b</sup>	748 $\pm$ 11.1 <sup>b</sup>	497 $\pm$ 12.3 <sup>c</sup>

Mean values within a row not sharing a common superscript letters (a, b, c, d) were significantly different,  $p < 0.05$ .

## DISCUSSION

The present study investigated the toxicity involved in TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination induced sub-chronic toxicity in male rats. The used rats exposure route to nanoparticles is the oral pathway because most of these particles are used in food packaging and so may gain entry into the body through this route. These nanoparticles are also used in other consumer products applications such as biomedical materials, diagnostic, imaging, therapeutic and drug delivery and thus there is always a risk of ingestion during use. Those doses and exposure route used do not necessarily reflect the concentration of nanoparticles used in the environment these doses and route were selected to study sub-chronic toxicity through oral exposure. There are several studies that suggest the toxicity of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs used in different dosages and exposure methods. Nanoparticles were found to reach the systemic circulation and disseminate to several organs such as the liver, spleen, brain, and kidney (Oberdörster et al., 2005 and Heringa et al., 2016). Nanoparticles can be induced disrupting cellular processes and causing disease penetrates lung, liver, brain, dermal barriers and enters the circulatory and lymphatic systems of humans and animals, reaching most bodily tissues and organs (Nel et al., 2006). Due to the small size of nanoparticles upon the particular arrangement of its many atoms, Nanoparticles can enter the blood circulation and lymphatic systems, and eventually into the tissues of the body and organs (Donaldson et al., 2005). Studies

dealing with oral exposure of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs in male rats demonstrated the presence of particles in distant organs. Biodistribution experiment showed that TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs could be transported to other tissues and organs through the blood circulation in rats after uptake by gastrointestinal tract including liver, spleen, kidney, and lung (Aust et al., 2002 and Wang et al., 2007).

In our previous study, we found oral administration with TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination for 60 consecutive days induced damage, inflammation, and biochemical alterations in plasma, liver and brain rats. To understand the role of inflammation in TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs oral exposure, the measurements show increased of inflammatory response like IFN $\gamma$ , TNF- $\alpha$  and IL-6 there were observed in liver and brain (Aust et al., 2002 and Zhang et al., 2010). Although there is growing concern in nanotoxicity studies, there is little known about the interaction of nanoparticles with the biological system. Furthermore, it has been demonstrated that TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs stimulate the release proinflammatory cytokines such as IFN $\gamma$ , TNF- $\alpha$  and IL-6 which play a major role in liver and brain inflammation by induced increase in the levels of cytokines and the effects was more pronounced in the combination group. Data show the effects of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs and their combination in liver and brain by observing the oxidative stress in the displayed results. Our results indicate an increased elevated NO levels in all the nanoparticles the combination was much higher than the other groups. Li et al. (2008) reported that nanoparticles induced toxic effects through the generation of various deleterious free radicals including, ROS like hydrogen peroxide, hydroxyl radical species, nitric oxide or superoxide anion. Elevated level of NO in liver and brain following exposure to TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination showed enhanced lipid peroxidation and free radicals. Several studies have indicated generation free radical in lipid peroxidation due to induced toxicity by nanoparticles (Li et al., 2008; Park et al., 2008 and Xia et al., 2006).

Further, a significant decrease in TAC level in TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs exposed group shows the reaction against the toxic effect of nanoparticles. The combination of TiO<sub>2</sub>NPs with CeO<sub>2</sub>NPs was more toxic than each TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs alone. Antioxidants play an important role in preventing, or in most cases, limiting the damage caused by ROS. The hydroxyl radical possesses the highest one electron reduction potential of all the physiologically relevant ROS, and is extremely reactive with almost every type of biomolecule, including proteins and nucleic acids (Wang et al., 2008 and Park et al., 2010). Furthermore, Valko et al. (2007) reported that all cells in the body contain powerful antioxidant enzymes. In addition, there are numerous specialized antioxidant enzymes reacting with and detoxifying oxidants. Neurotransmitters are play a role in memory, awareness, thought, and consciousness and allow the organism to become alert and guards against the intensification of reflex reactions and other behavior. Few studies that have indicated nanoparticles can cross the blood-brain barrier and enter the central nervous system (CNS). Moreover, impaired glutamate and ACh activities in TiO<sub>2</sub>NPs nanoparticles exposed animals indicating disruption in the normal functioning of CNS (Sharma & Sharma, 2007; Wang et al., 2008 and Ma et al., 2010). It is noted in recent research that serotonin which acts as a neurotransmitter and modulate gastrointestinal motility, peripheral vascular tone, cerebral vascular tone, and platelet function and has been implicated in the pathophysiology of mood disorders, emesis, migraine and irritable bowel syndrome (IBS) (Mohammad-Zadeh et al., 2008). Studies have shown the accumulation of NPs in brain caused in changed synthesis and release of certain neurotransmitters and receptors in nerve cells, leading to brain damage in rats (Ma et al., 2010).

(DA) is neurotransmitters, which is found in neurons of all animals. Alteration in the normal expression of this transmitter is associated with human neurological disorders such as Parkinson's disease and depression. Moreover, nanoparticles led to an increased level of DA in cortex region of mouse brain suggesting weakened ability to maintain an appropriate state of activation in the CNS due to toxicity. As well excessive production of ROS may lead to selective toxicity to DA neurons (Deakin, 2003 and Yoo et al., 2003). The study was considered to the potential role of toxic TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination on the brain. Therefore, our data show that treatment with TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination caused decreased in DA and serotonin, while Ach were increased in plasma and brain compared to control group. In the combination group the effect was more pronounced than each group.

Liver is the major organ for biotransformation of toxins and as a result; it may be the first organ to be exposed to nanoparticles that are able to enter into the circulation. Oral exposure of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs in rats demonstrated the presence of nanoparticles in distant organs. As a result nanoparticles can be transported to other tissues and organs through the blood circulation in rats after uptake by gastrointestinal tract including liver, brain, spleen, kidney, and lung (Wang et al., 2007; Nalabotu et al., 2011 and Tassinari et al., 2014). The accumulation of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs in liver causing toxicity revealed by disruption of liver function enzymes including AST, ALT, AIP and LDH. As well as induced cell death occurs through oxidative stress and DNA fragmentation in the hepatic cells (Wang et al., 2007; Geraets et al., 2012 and Mohammad et al., 2017). Data showed that treatment with TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs and their combination caused increase plasma and decrease liver ALT, AST, AIP and LDH activities compared to control group. Co-treatment of TiO<sub>2</sub>NPs plus CeO<sub>2</sub>NPs was more effect than each other. Several studies have indicated that high level of AST activity can suggest a large proportion is hepatocellular injury. ALT is found in many organs and catalyzes the transfer



of the  $\alpha$ -amino group from alanine to  $\alpha$ -ketoglutaric acid and that the level of height will be special in cytoplasmic liver. There are many enzymes that will be excreted when there is damage to the hepatic cell membrane including AST, ALT, AIP, AcP and LDH are secreted into the blood (Ncibi et al., 2008 and Park et al., 2009).

## CONCLUSION

The present study showed that of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs alone or their combination induced hepatotoxicity and neurotoxicity in liver and brain. They changed in tissues inflammatory markers (IFN $\gamma$ , TNF- $\alpha$  and IL-6). Also, (TAC) was decreased in liver and brain tissues while (NO) was increased. Moreover, decreased (DA) and serotonin in plasma and brain while (ACh) was increased compared to control group. Also, (ALT, AST, AIP and LDH) were increased in plasma and decreased in liver. The hepatotoxicity and neurotoxicity was more pronounced in co-treatment of TiO<sub>2</sub>NPs with CeO<sub>2</sub>NPs than each other one. Therefore, Nanoparticles have a potential toxicity when exposed, so it is necessary to evaluate the resulting risks to ensure the safe use and disposal of nanoparticles.

## SUMMARY OF RESEARCH

TiO<sub>2</sub>NPs are usually used in wide applications due to its chemical and physical characteristics, air purification in biomedical fields, cosmetics industry, sewage purification, food packaging and painting. Also, CeO<sub>2</sub>NPs it is widely used applications as a catalyst, ultraviolet absorber and biomedical materials because of the redox activities, can be used as reducing fuel consumption, greenhouse gases and particle numbers in vehicle exhaust, additive to improve the burning efficiency of fuels. Thus, the environmental and health effect of TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs are of great interest. Though TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs have a variety of applications, few studies have demonstrated that exposure to TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs may lead to adverse effects, such as hepatotoxicity and neurotoxicity. The previous studies demonstrated that TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs induced oxidative stress and lipid peroxidation in liver and brain. Therefore, the present study aimed to investigate the hepatotoxicity and neurotoxicity of TiO<sub>2</sub>NPs, CeO<sub>2</sub>NPs and their combination in male rats.

## FUTURE ISSUES

Must be reduce the exposure to TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs alone or in combination which used in a variety of application such as drug delivery, industrial and healthcare applications, biomedical fields, cosmetics industry, sewage purification, food packaging, painting, catalyst, ultraviolet absorber and biomedical materials. Additional studies are needed to demonstrate the hepatotoxicity and neurotoxicity induced by different doses, sizes and time of exposure to TiO<sub>2</sub>NPs and CeO<sub>2</sub>NPs alone or in combination.

## Acknowledgment

I would like to express my thanks and great gratitude to all the brothers in the analysis laboratories in Cairo and Alexandria who helped me to complete this manuscript.

## Funding:

This study has not received any external funding.

## Conflict of Interest:

The authors declare that there are no conflicts of interests.

## Data and materials availability:

All data associated with this study are present in the paper.

## REFERENCE

1. Aalapathi S, Ganapathy S, Manapuram S, Anumolu G, Prakya BM. Toxicity and bio-accumulation of inhaled cerium oxide nanoparticles in CD1 mice. *Nanotoxicol.* 2014; 8, 786-798.
2. Aust AE, Ball JC, Hu AA, Lighty JS, Smith KR, Straccia AM, Veranth JM, Young WC. Particle characteristics responsible for effects on human lung epithelial cells. Research Report (Health Effects Institute). 2002, 110, 1-65.

3. De Jong WH, Borm PJ. Drug delivery and nanoparticles: applications and hazards. *Int.J.Nanomed.* 2008, 3, 133.
4. Deakin JF. Depression and antisocial personality disorder: two contrasting disorders of 5HT function. In *Neuropsychopharmacology 2003* (pp. 79-93). Springer, Vienna.
5. Donaldson K, Tran L, Jimenez LA, Duffin R, Newby DE, Mills N, MacNee W, Stone V. Combustion-derived nanoparticles: a review of their toxicology following inhalation exposure. *Part.Fib. Toxicol.* 2005, 2, 10.
6. Duncan DB. Multiple range and multiple F tests. *Biometrics.* 1955, 11, 1-42.
7. Ferguson-Smith AC, Chen YF, Newman MS, May LT, Sehgal PB, Ruddle FH. Regional localization of the interferon- $\beta$ 2B-cell stimulatory factor 2/hepatocyte stimulating factor gene to human chromosome 7p15-p21. *Genomics.* 1988, 2, 20320-8.
8. Geraets L, Oomen AG, Schroeter JD, Coleman VA, Cassee FR. Tissue distribution of inhaled micro- and nano-sized cerium oxide particles in rats: results from a 28-day exposure study. *Toxicol. Sci.* 2012, 127, 463-473.
9. Gurr JR, Wang AS, Chen CH, Jan KY. Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicol.* 2005, 213, 66-73.
10. Hedayati M, Yazdanparast R, Azizi F. Determination of human tumor necrosis factor  $\alpha$  by a highly sensitive enzyme immunoassay. *Biochem. Biophys. Res. Commun.* 2001, 289, 295-298.
11. Heringa MB, Geraets L, van Eijkeren JC, Vandebriel RJ, de Jong WH, Oomen AG. Risk assessment of titanium dioxide nanoparticles via oral exposure, including toxicokinetic considerations. *Nanotoxicol.* 2016, 10, 1515-1525.
12. Hirst SM, Karakoti A, Singh S, Self W, Tyler R, Seal S, Reilly CM. Bio-distribution and in vivo antioxidant effects of cerium oxide nanoparticles in mice. *Environ. Toxicol.* 2013, 28, 107-118.
13. Jaeger A, Weiss DG, Jonas L, Kriehuber R. Oxidative stress-induced cytotoxic and genotoxic effects of nano-sized titanium dioxide particles in human HaCaT keratinocytes. *Toxicol.* 2012, 296, 27-36.
14. Kumari M, Kumari SI, Kamal SS, Grover P. Genotoxicity assessment of cerium oxide nanoparticles in female Wistar rats after acute oral exposure. *Mut. Res/Gen. Toxicol. Environ. Mutagen.* 2014, 31, 7-19.
15. Lee SS, Song W, Cho M, Puppala HL, Nguyen P, Zhu H, Segatori L, Colvin VL. Antioxidant properties of cerium oxide nanocrystals as a function of nanocrystal diameter and surface coating. *ACS Nano.* 2013, 7, 9693-9703.
16. Li SQ, Zhu RR, Zhu H, Xue M, Sun XY, Yao SD, Wang SL. Nanotoxicity of TiO<sub>2</sub> nanoparticles to erythrocyte *in vitro*. *Food. Chem. Toxicol.* 2008, 46, 3626-3631.
17. Lin W, Stayton I, Huang YW, Zhou XD, Ma Y. Cytotoxicity and cell membrane depolarization induced by aluminum oxide nanoparticles in human lung epithelial cells A549. *Toxicol. Environ. Chem.* 2008, 90, 983-996.
18. Ma JY, Mercer RR, Barger M, Schwegler-Berry D, Scabilloni J, Ma JK, Castranova V. Induction of pulmonary fibrosis by cerium oxide nanoparticles. *Toxicol. Appl. Pharmacol.* 2012, 262, 255-264.
19. Ma L, Liu J, Li N, Wang J, Duan Y, Yan J, Liu H, Wang H, Hong F. Oxidative stress in the brain of mice caused by translocated nanoparticulate TiO<sub>2</sub> delivered to the abdominal cavity. *Biomateri.* 2010, 31, 99-105.
20. Mohammad F, Arfin T, Al-Lohedan HA. Enhanced biological activity and biosorption performance of trimethyl chitosan-loaded cerium oxide particles. *J. Ind. Eng. Chem.* 2017, 45, 33-43.
21. Mohammad-Zadeh LF, Moses L, Gwaltney-Brant SM. Serotonin: a review. *J. Veterin. Pharmacol. Ther.* 2008, 31, 187-99.
22. Morgan AM, Ibrahim MA, Noshay PA. Reproductive toxicity provoked by titanium dioxide nanoparticles and the ameliorative role of Tiron in adult male rats. *Biochem. Biophys. Res. Commun.* 2017, 486, 595-600.
23. Nalabotu SK, Kolli MB, Triest WE, Ma JY, Manne ND, Katta A, Addagarla HS, Rice KM, Blough ER. Intratracheal instillation of cerium oxide nanoparticles induces hepatic toxicity in male Sprague-Dawley rats. *Int. J. Nanomed.* 2011, 6, 2327.
24. Ncibi S, Othman MB, Akacha A, Krifi MN, Zourgui L. Opuntia ficus indica extract protects against chlorpyrifos-induced damage on mice liver. *Food. Chem. Toxicol.* 2008, 46, 797-802.
25. Nel A, Xia T, Mädler L, Li N. Toxic potential of materials at the nanolevel. *science.* 2006, 11, 622-627.
26. (NRC) National Research Council. Guide for the care and use of laboratory animals. Institute of Laboratory Animal Resources, Commission on Life Sciences. National Academy of Sciences, Washington, DC. 1996.
27. Oberdörster G, Maynard A, Donaldson K, Castranova V, Fitzpatrick J, Ausman K, Carter J, Karn B, Kreyling W, Lai D, Olin S. Principles for characterizing the potential human health effects from exposure to nanomaterials: elements of a screening strategy. *Part.Fib. Toxicol.* 2005, 2, 8.
28. Park EJ, Cho WS, Jeong J, Yi JH, Choi K, Kim Y, Park K. Induction of inflammatory responses in mice treated with cerium oxide nanoparticles by intratracheal instillation. *J. Health. Sci.* 2010, 56, 387-396.



29. Park EJ, Yi J, Chung KH, Ryu DY, Choi J, Park K. Oxidative stress and apoptosis induced by titanium dioxide nanoparticles in cultured BEAS-2B cells. *Toxicol. Lett.* 2008, 180, 222-229.
30. Park EJ, Yoon J, Choi K, Yi J, Park K. Induction of chronic inflammation in mice treated with titanium dioxide nanoparticles by intratracheal instillation. *Toxicol.* 2009, 260, 37-46.
31. Paur HR, Cassee FR, Teeguarden J, Fissan H, Diabate S, Aufderheide M, Kreyling WG, Hänninen O, Kasper G, Riediker M, Rothen-Rutishauser B. In-vitro cell exposure studies for the assessment of nanoparticle toxicity in the lung-A dialog between aerosol science and biology. *J. Aerosol Sci.* 2011, 42, 668-692.
32. Sadiq R, Bhalli JA, Yan J, Woodruff RS, Pearce MG, Li Y, Mustafa T, Watanabe F, Pack LM, Biris AS, Khan QM. Genotoxicity of TiO<sub>2</sub>anatase nanoparticles in B6C3F1 male mice evaluated using Pig-a and flow cytometric micronucleus assays. *Muta. Res/Genet. Toxicol. Environ Mutagen.* 2012, 745, 65-72.
33. Sarkar A, Ghosh M, Sil PC. Nanotoxicity: oxidative stress mediated toxicity of metal and metal oxide nanoparticles. *J. Nanosci. Nanotech.* 2014, 14, 730-743.
34. SAS, Statistical Analysis System. SAS Procedure Guide. Release 6.03 Edition. SAS Institute Inc., Cary, Nc, U.S.A 1998.
35. Selvan VA, Anand RB, Udayakumar M. Effects of cerium oxide nanoparticle addition in diesel and diesel-biodiesel-ethanol blends on the performance and emission characteristics of a CI engine. *J EngAppl Sci.* 2009, 4, 1819-6608.
36. Shalaby MR, Aggarwal BB, Rinderknecht E, Svedersky LP, Finkle BS, Palladino MA. Activation of human polymorphonuclear neutrophil functions by interferon-gamma and tumor necrosis factors. *J. Immunol.* 1985, 135, 2069-2073.
37. Sharma HS, Sharma A. Nanoparticles aggravate heat stress induced cognitive deficits, blood-brain barrier disruption, edema formation and brain pathology. *Prog.Brain. Res.* 2007, 162, 245-273.
38. Shi H, Magaye R, Castranova V, Zhao J. Titanium dioxide nanoparticles: a review of current toxicological data. *Part.FibreToxicol.* 2013, 10, 15.
39. Srinivas A, Rao PJ, Selvam G, Murthy PB, Reddy PN. Acute inhalation toxicity of cerium oxide nanoparticles in rats. *Toxicol Lett.* 2011, 205, 105-115.
40. Tassinari R, Cubadda F, Moracci G, Aureli F, D'Amato M, Valeri M, De Berardis B, Raggi A, Mantovani A, Passeri D, Rossi M. Oral, short-term exposure to titanium dioxide nanoparticles in Sprague-Dawley rat: focus on reproductive and endocrine systems and spleen. *Nanotoxicol.* 2014, 8, 654-662.
41. Tseng MT, Lu X, Duan X, Hardas SS, Sultana R, Wu P, Unrine JM, Graham U, Butterfield DA, Grulke EA, Yokel RA. Alteration of hepatic structure and oxidative stress induced by intravenous nanoceria. *Toxicol.Appl. Pharmacol.* 2012, 260, 173-182.
42. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem.Cell. Biol.* 2007, 39, 44-84.
43. Wang CH, Lin SS. Preparing an active cerium oxide catalyst for the catalytic incineration of aromatic hydrocarbons. *Appl.Catal. A: General.* 2004, 268, 227-233.
44. Wang J, Li N, Zheng L, Wang S, Wang Y, Zhao X, Duan Y, Cui Y, Zhou M, Cai J, Gong S. P38-Nrf-2 signaling pathway of oxidative stress in mice caused by nanoparticulate TiO<sub>2</sub>. *Biol. Trace Elem. Res.* 2011, 140, 186-197.
45. Wang J, Liu Y, Jiao F, Lao F, Li W, Gu Y, Li Y, Ge C, Zhou G, Li B, Zhao Y. Time-dependent translocation and potential impairment on central nervous system by intranasally instilled TiO<sub>2</sub> nanoparticles. *Toxicol.* 2008, 254, 82-90.
46. Wang J, Zhou G, Chen C, Yu H, Wang T, Ma Y, Jia G, Gao Y, Li B, Sun J, Li Y. Acute toxicity and biodistribution of different sized titanium dioxide particles in mice after oral administration. *Toxicol. Lett.* 2007, 168, 176-185.
47. Wu J, Liu W, Xue C, Zhou S, Lan F, Bi L, Xu H, Yang X, Zeng FD. Toxicity and penetration of TiO<sub>2</sub> nanoparticles in hairless mice and porcine skin after subchronic dermal exposure. *Toxicol. Lett.* 2009, 191, 1-8.
48. Xia T, Kovichich M, Nel A. The role of reactive oxygen species and oxidative stress in mediating particulate matter injury. *ClinOccup Environ Med.* 2006, 5, 817-836.
49. Xu C, Qu X. Cerium oxide nanoparticle: a remarkably versatile rare earth nanomaterial for biological applications. *NPG Asia Materials.* 2014, 6, e90.
50. Yoo MS, Chun HS, Son JJ, DeGiorgio LA, Kim DJ, Peng C, Son JH. Oxidative stress regulated genes in nigral dopaminergic neuronal cells: correlation with the known pathology in Parkinson's disease. *Mol. Brain Res.* 2003, 110, 76-84.
51. Yu Y, Ren W, Ren B. Nanosize titanium dioxide cause neuronal apoptosis: a potential linkage between nanoparticle exposure and neural disorder. *Neurol.Res.* 2008, 30, 1115-1120.
52. Zhang R, Niu Y, Li Y, Zhao C, Song B, Li Y, Zhou Y. Acute toxicity study of the interaction between titanium dioxide nanoparticles and lead acetate in mice. *Environ. ToxicolPharmacol.* 2010, 30, 52-60.